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The reaction of 2,3-butanedione, ethyl pyruvate, and phenylglyoxal with β -nitrostyrene and L-proline in isopropanol at room temperature gives substituted pyrrolizidines, as a result of one-pot three component reaction. On the contrary, a spiropyrrolizidine is formed from 1,2-cyclohexanedione only when the reaction is carried out in refluxing isopropanol, whereas at room temperature, incorporation of the amine component into the products is not observed and bicyclo[3.2.1]octanones are formed, as a result of a tandem Michael-Henry reaction. In this latter case, L-proline acts as an organocatalyst, although with modest enantioselectivity. The stereochemistry of the products is given and the mechanism of formation of products is postulated, on the basis of stereochemical arguments.

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INTRODUCTION

The 1-azabicyclo[3.3.0]octane ring is part of naturally occurring and biologically active pyrrolizidine alkaloids, which are studied for their interesting diverse physiological properties [1]. Construction, also asymmetric, of the pyrrolizidine ring are usually made through multistep reactions [2] and also [3+2]cycloadditions between 1,3azomethine ylides and activated olefins are widely used [3]. A possible source of 1,3-azomethine ylides is derived from activated carbonyl compounds and L-proline, which can loose carbon dioxide when heated. The resulting 1,3-dipoles can react with aldehydes, to give bicyclic 1,3-oxazolidines [4], or with α , β -unsaturated esters [3(a,e,f)], or nitroolefins [3(b)] to give pyrrolizidine derivatives.

In this work, we report the application of 1,3-azomethine ylides derived from α -dicarbonyl compounds and L-proline to the synthesis of pyrrolizidines by means of 1,3-dipolar cycloaddition reactions to β -nitrostyrene. The use of (2*S*,4*R*)-4-hydroxyproline gave enantiopure polysubstituted pyrrolizidines.

RESULTS AND DISCUSSION

The α -dicarbonyl compounds examined in this study have been 2,3-butanedione 1a, ethyl pyruvate 1b, phenylglyoxal 1c (Scheme 1), and 1,2-cyclohexanedione 9 (Scheme 3), which were reacted with β -nitrostyrene 2 in isopropanol at room temperature, in the presence of an equimolar amount of L-proline. The reaction of 2,3-butanedione 1a with β -nitrostyrene 2 in isopropanol at room temperature for 3 days gave two regioisomeric cycloadducts in 3:2 ratio, which were assigned the structures 3a and 4a, respectively, on the basis of difference NOE measurements (Table 1). The analogous cycloaddition of ethyl pyruvate 1b to 2 gave two adducts 3b and 4b in 9:1 ratio (Scheme 1), with structural assignments based on analogy and difference NOE measurements (Table 1), while the addition of phenylglyoxal 1c to 2 furnished a single regioisomer 3c.

To determine the mechanism of formation of the pyrrolizidines, a reaction was carried out between 2,3-butanedione **1a**, β -nitrostyrene **2** and (2S,4R)-4-hydroxyproline **5** in isopropanol, so to take advantage of the

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presence of a stereocenter of known configuration. The reaction was much slower, owing to solubility problems, and gentle warming was necessary to drive the reaction to completion. The reaction products isolated were the pyrrolizidines **6a** and **7a**, analogous to **3a** and **4a**, together with a third isomer **8a**, whose formation can be surely ascribed to the higher temperature used. The relative ratio of compounds **6a–8a** was 1:4:1 (Scheme 2).

X-ray analysis of compound 6a (Fig. 1) showed the geometry of the substituents and the *cis* fusion of the rings. To validate the use of NOE measurements to determine the geometry of the other pyrrolizidines, difference NOE experiments (Table 1) were carried out also on 6a, whose results were in complete accordance with the X-ray determination. The ring junction in all pyrrolizidines was determined by an analysis of their IR

spectra. The Bohlmann bands [5] between 2800 and 2600 cm⁻¹, which are correlated with the presence of a C—H bond antiperiplanar to the nitrogen lone pair, were present only for compound **8a** (2724 and 2671 cm⁻¹), which was therefore assigned the *trans* fusion, whereas pyrrolizidines **3a–c**, **4a,b** and **6a**, **7a** which lack these bands, were assigned the *cis* fusion.

1,2-Cyclohexanedione **9** has been chosen as another candidate for synthesizing pyrrolizidine target compounds. Indeed, the reaction of the substrate **9** with β -nitrostyrene **2**, carried out in refluxing isopropanol for 2 h gave as the main reaction product (90%) the nitrospiro pyrrolizidine **10** (Scheme 3, right side). Its stereochemistry was assigned as depicted in Scheme 3. The relative configuration of the spiro carbon atom C-3' was confirmed by the resonance value found for the axial proton

Difference from experiments carried out on the pyrionizations.						
	Irradiated protons					
Compound	H-1 (H-1' for 10)	H-2 (H-2' for 10)	H-3	Н-6	H-7a (H-7'a for 10)	Me
3a	o-Ph (8), H-7a (9)	Me (4), H-5 at 2.91 (6), H-7 at 1.33 (2)			H-1 (9), H-7 at 2.04 (5)	H-2 (4)
3b	o-Ph (10), H-7a (10)	Me (9), H-5 at 2.90 (4)			H-1 (10), H-7 at 2.07 (6)	H-2 (9)
3c	o-Ph (7), H-7a (11)	H-3 (7), H-5 at 2.81 (4)	H-2 (9), H-5		H-1 (10), o-Ph (7),	
			at 2.81 (6)		H-7 at 2.06 (8)	
4 a	Me (6), H-5 at 3.19 (5),	o-Ph (5), H-7a (3)			o-Ph (6), H-2 (3),	H-1 (6), H-5
	H-7 at 1.75 (4)				MeCO (2)	at 3.19 (5)
4b	H-5 at 3.13 (3), H-7	o-Ph (8), H-7a (4)			Me (4), <i>o</i> -Ph (11),	H-7a (4), H-5
	at 1.72 (4), Me (3)				H-2 (4),	at 3.13 (3)
					H-5 at 3.13 (3)	
6a	o-Ph (11), H-7a (8)	Me (7), H-5 at 3.12 (5),		H-2 (4), H-5	H-1 (8), H-7	H-2 (4)
		H-6 (4)		at 3.12 (3),	at 2.15 (3)	
				H-7 at 1.63 (4)		
7a	H-5 at 3.37 (6), H-7	o-Ph (5), H-7a (5),		H-5 at 3.37 (5),	o-Ph (4), MeCO (2),	H-5 at 3.37 (2),
	at 2.18 (6)	H-7 at 2.03 (4)		H-7 at 2.18 (8)	H-7 at 2.03 (3)	MeCO (2)
8a	H-5 at 3.21 (3), H-7	<i>o</i> -Ph (8)		H-5 at 3.34 (5),	<i>o</i> -Ph (5)	H-5 at 3.21 (6)
	at 1.74 (6), Me (1)			H-7 at 2.32 (4)	· ·	
10	H-7 at 3.20 (2)	o-Ph (7), H-5 at 1.64 (2)			<i>o</i> -Ph (5)	

 Table 1

 Difference NOE experiments carried out on the pyrrolizidines

 η Values (%) in parenthesis for enhanced protons.



at C-2 which is strongly deshielded with respect to its geminal equatorial one (3.20 vs. 2.39 ppm) by the anisotropic effect of the nitro group that is spatially close to it.

Interestingly, when the same reaction was carried out at room temperature, even under solvent-free conditions, two adducts were obtained in 9:1 ratio (Scheme 3, left side). These compounds were assigned the structures **11** and **12**, on the basis of ¹H NMR spectra and X-ray analysis carried out on compound **12**. The two isomers were separated by flash chromatography and they proved weakly optically active $([\alpha]_D^{25} = +0.05 \ (c \ 2.65, MeOH) \ for$ **11** $and <math>[\alpha]_D^{25} = -$ 2.6 (c 0.15, MeOH) for **12**, 14% e.e., determined by HPLC on chiral column), thus demonstrating the organocatalytic capability of L-proline, albeit modest, in this particular case.

X-ray analysis of compound **12** (Fig. 2) showed that the geometry of phenyl and the nitro group was *cis*, both groups being *exo* oriented. The stereochemistry of its diastereomer **11** was determined by ¹H NMR analysis. In fact, in **12**, the coupling constant between H-6 and H-7 was larger than in **11** ($J_{6,7} = J_{endo,exo} = 5.8$ Hz for **11**, $J_{6,7} = J_{endo,endo} = 9.5$ Hz for **12**), thus demonstrating the *trans* relationship between the phenyl group



Figure 1. Molecular structure of compound 6a.

and the nitro group in **11** [6]. The orientation of the phenyl group was assigned as *exo* also in **11**. In fact, the coupling constant between H-5 and the benzylic proton H-6 was zero, thus indicating a dihedral angle of 90° between them, which is only consistent with the *endo* orientation of the benzylic proton itself.

MECHANISM OF THE REACTIONS

The mechanism of pyrrolizidine formation can be inferred from the stereochemistry of the products and in particular from compounds 6a and 7a, whose absolute configurations are known, given the R absolute configuration of the carbinol carbon atom. Since the majority of the pyrrolizidines in this work are cis fused [7], the first formed carbon-carbon bond (C1-C7a) occupies the same position as the carboxy group in the original pyrrolidine ring. Therefore, it seems reasonable to postulate that formation of the intermediate 13 (Scheme 4, illustrated for 2,3-butanedione 1a as the substrate) is immediately followed by elimination of carbon dioxide with the creation of the 1,3-dipole 14, existing in two resonance structures a and **b**. Both regioisomeric adducts **6a** and **7a** derived from the less encumbered exo-anti transition states, while adduct 8a arose from the exo-syn transition state. The same reaction mechanism may be invoked for the formation of the other pyrrolizidines 3a-c, 4a,b derived from linear 1,2,dicarbonyl compounds and 10 derived from 1,2cyclohexanedione.

In the reaction of 1,2-cyclohexanedione 9 with β -nitrostyrene 2 performed at room temperature, L-proline initially reacts with the substrate 9 to give the corresponding crossconjugated enaminone 15 (Scheme 5) which reacts with the nitroolefin in a Michael-type addition, through a two step mechanism, with subsequent ring closure in 16, by collapse of the carbanion onto the carbonyl carbon atom. The consequent zwitterion 17 is hydrolysed *in situ* to afford eventually the two diastereomers 11 and 12.

The easier loss of carbon dioxide observed for the adduct between L-proline and 2,3-butanedione with respect to the analogous intermediate from 1,2-cyclohex-anedione could be ascribed to the s-*trans* geometry

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Scheme 3



between the carbonyl group and the iminium group in the former which would facilitate the expulsion of carbon dioxide in the former compound.

EXPERIMENTAL

IR spectra were recorded on a Jasco FT/IR 200 spectrophotometer. ¹H NMR and ¹³C NMR spectra were run on a Jeol EX-400 spectrometer (400 MHz for proton, 100.1 MHz for carbon), and on a Jeol EX-270 spectrometer (270 MHz for proton, 68 MHz for carbon) using deuteriochloroform as the solvent and tetramethylsilane as the internal standard. Chemical shifts are expressed in parts per million (δ). Coupling constants are given in Hz. Optical rotations were determined on a Perkin Elmer Model 241 polarimeter. Mass spectra were recorded on a ion trap instrument Finnigan GCQ (70 eV). HPLC analysis were run on a Hewlett Packard Series 1100 instrument the chiral column being a Lux 5 µm Cellulose-2 (Phenomenex) with a Cellulose tris(3-chloro-4-methylphenylcarbammate) chiral stationary phase, eluent: n-hexane/isopropanol 75:25, detector UV 220 nm; TLC's were performed on Polygram[®] Sil G/UV₂₅₄ silica gel pre-coated plastic sheets. Flash chromatography was run on silica gel for flash-chromatography (BDH). Elemental analyses were determined on a Carlo Erba 1106 instrument, at the Department of Chemical Sciences and Technologies of the University of Udine, Italy. Light petroleum refers to the fraction with b.p. 40–70°C and ether to diethyl ether. 2,3-Butanedione, L-proline, (2S,4R)-4-



Figure 2. Molecular structure of compound 12.

hydroxyproline, and \beta-nitrostyrene were purchased from Sigma-Aldrich and 1,2-cyclohexanedione was purchased from Lancaster.

General procedure for the reactions between α -dicarbonyl compounds 1a-c and 9, β -nitrostyrene 2 and L-proline or (2S,4R)-4-hydroxyproline 5. To the α -dicarbonyl compound (1.0 mmol) in isopropanol (10 mL), \beta-nitrostyrene (1.0 mmol) and the appropriate proline derivative (1.0 mmol) were added. The mixture was set aside at room temperature until completion of the reaction, which needed different times depending on the substrate (96 h for diacetyl and 24 h for 1,2cyclohexanedione). The solvent was eliminated and the crude reaction mixture was chromatographed on silica gel (eluent: nheptane-ethyl acetate, gradient).

(1S*,2S*,3R*,7aS*)-3-Acetyl-3-methyl-1-nitro-2-phenylazabicyclo[3.3.0]octane (3a). Oil, 52% yield, after purification. $R_{\rm f}$ 0.15 (eluent: light petroleum–ethyl acetate 4:1). IR (neat) 3050, 3032, 1603, 1498, 761, 733, 702 (Ph), 1708 (C=O), 1545, 1375, 1352 (NO₂); ¹H NMR (400 MHz, CDCl₃): δ 7.28 (m, 5H, Ph), 5.95 (dd, $J_1 = 10.6$ Hz, $J_2 = 9.1$ Hz, 1H, H-1), 4.30 (dt, $J_1 = 9.1$ Hz, $J_2 = J_3 = 7.8$ Hz, 1H, H-7a), 3.85 (d, J = 10.6 Hz, 1H, H-2), 3.04 (m, 1H, H-5), 2.93 (ddd, $J_1 = 11.2$ Hz, $J_2 = 8.1$ Hz, $J_3 = 5.3$ Hz, 1H, H-5), 2.05 (m, 1H, H-7), 1.94 (m, 1H, H-6), 1.91 (s, 3H, CH₃CO), 1.83 (m, 1H, H-6), 1.38 (s, 3H, CH₃), 1.33 (m, 1H, H-7); ¹³C NMR (100.1 MHz, CDCl₃): δ 207.7 (s), 133.8 (s), 129.0 (2d), 128.6 (2d), 127.7 (d), 93.1 (d, C-1), 74.5 (s, C-3), 64.0 (d, C-7a), 55.2 (d, C-2), 48.9 (t, C-5), 27.7 (t, C-7), 27.1 (q, CH₃CO), 25.6 (t, C-6), 19.2 (q, CH₃); *m*/*z* 289 (2, M+1), 245 (45, M+1—CH₃CO), 199 (100, 245-NO₂), 184 (56, 199-CH₃). Anal. Calcd. for $C_{16}H_{20}N_2O_3$: C, 66.65; H, 6.99; N, 9.72; Found: C, 66.82; H, 7.12; N, 9.88.

(1S*,2R*,3R*,7aS*)-3-Acetyl-3-methyl-2-nitro-1-phenylazabicyclo[3.3.0]octane (4a). Oil, 35% yield, after purification. $R_{\rm f}$ 0.65 (eluent: light petroleum–ethyl acetate 4:1). IR (neat) 3063, 3030, 1603, 1496, 742, 700 (Ph), 1714 (C=O), 1548, 1361 (NO₂); ¹H NMR (400 MHz, CDCl₃): δ 7.36 (m, 3H, Ph), 7.25 (bd, 2H, Ph), 6.09 (d, J = 8.4 Hz, 1H, H-2), 3.84 (dd, $J_1 = 8.4$ Hz, $J_2 = 9.6$ Hz, 1H, H-1), 3.22 (m, 1H, H-7a), 3.19 (dd, $J_1 =$ 8.8 Hz, $J_2 = 6.9$ Hz, 1H, H-5), 3.01 (m, 1H, H-5), 2.46 (s, 3H, CH₃CO), 2.10 (m, 1H, H-6), 2.05 (m, 2H, H-6, H-7), 1.76 (m, 1H, H-7), 1.46 (s, 3H, CH₃); ¹³C NMR (100.1 MHz, CDCl₃): δ 207.6 (s), 137.8 (s), 128.9 (2d), 127.8 (2d), 127.5 (d), 98.4 (d, C-2), 76.4 (s, C-3), 70.8 (d, C-7a), 56.7 (d, C-1), 46.4 (t, C-5), 28.2 (t, C-7), 24.4 (t, C-6), 24.1 (q, CH₃CO), 16.1 (q, CH₃); m/z 289 (5, M+1), 245 (40, M+1-CH₃CO), 199 (100, 245-NO₂), 184 (60, 199-CH₃). Anal. Calcd. for C₁₆H₂₀N₂O₃: C, 66.65; H, 6.99; N, 9.72; Found: C, 66.91; H, 6.80; N, 9.56.



(1S*,2S*,3R*,7aS*)-3-Methyl-1-nitro-2-phenylazabicyclo[3. 3.0] octane-3-carboxylic acid ethyl ester (3b). Mp 101-102°C, 85% yield, after purification. Rf 0.30 (eluent: n-heptane-ethyl acetate 4:1). IR (CHCl₃) 3080, 3060, 1598, 1498, 770, 701, 669 (Ph), 1686 (C=O), 1549, 1376 (NO₂); ¹H NMR (400 MHz, CDCl₃): δ 7.28 (m, 3H, Ph), 7.19 (m, 2H, *o*-Ph), 5.90 (dd, $J_1 = 10.6$ Hz, $J_2 = 9.1$ Hz, 1H, H-1), 4.64 (bq, J =9.1 Hz, 1H, H-7a), 3.99 (dq, 2H, CH_2O), 3.90 (d, J = 10.6Hz, 1H, H-2), 3.04 (m, 1H, H-5 cis to H-7a), 2.90 (m, 1H, H-5 trans to H-7a), 2.07 (m, 1H, H-7 cis to H-7a), 1.93 (m, 1H, H-6), 1.85 (m, 1H, H-6), 1.48 (s, 3H, CH₃), 1.35 (m, 1H, H-7), 1.08 (t, 3H, CH₃CH₂O); ¹³C NMR (100.1 MHz, CDCl₃): δ 172.6 (s), 133.7 (s), 128.6 (2d), 128.1 (2d), 128.0 (d), 93.3 (d, C-1), 72.4 (s, C-3), 65.0 (d, C-7a), 61.0 (t, CH₂O), 55.8 (d, C-2), 49.5 (t, C-5), 27.9 (t, C-7), 25.5 (t, C-6), 18.5 (q, CH₃), 13.8 (q, CH₃CH₂O); *m*/*z* 319 (12, M+1), 272 (38, M—NO₂), 245 (50, M-COOEt), 199 (30, M-COOEt-NO2), 198 (100, M-COOEt-HNO₂), 183 (15), 170 (20). Anal. Calcd. for C17H22N2O4: C, 64.13; H, 6.97; N, 8.80; Found: C, 63.85; H, 6.83; N, 8.73.

(1S*,2R*,3R*,7aS*)-3-Methyl-2-nitro-1-phenylazabicyclo[3.3.0] octane-3-carboxylic acid ethyl ester (4b). Oil, 5% yield, after purification. R_f 0.50 (eluent: *n*-heptane–ethyl acetate 4:1). IR (neat) 3064, 3031, 1603, 1498, 752, 701 (Ph), 1723 (C=O), 1550, 1367 (NO₂); ¹H NMR (400 MHz, CDCl₃): δ 7.30 (m, 5H, Ph), 5.92 (d, J = 9.6 Hz, 1H, H-2), 4.30 (dq, 2H, CH₂O), 3.72 (q, 1H, H-7a), 3.69 (dd, $J_1 = 8.4$ Hz, $J_2 = 9.6$ Hz, 1H, H-1), 3.13 (dd, $J_1 = 9.0$ Hz, $J_2 = 6.8$ Hz, 1H, H-5 *trans* to H-7a), 2.93 (m, 1H, H-5 *cis* to H-7a), 2.00 (3H, m, 2 H-6, H-7), 1.76 (m, 1H, H-7), 1.45 (s, 3H, CH₃), 1.34 (t, 3H, CH₃CH₂O); ¹H NMR (400 MHz, CD₃OH): δ 7.30 (m, 5H, Ph), 5.90 (d, J = 9.9 Hz, 1H, H-2), 4.29 (dq, 2H, CH₂O), 3.71 (t, J₁ = 9.9 Hz, J₂ = 10.1 Hz, 1H, H-1), 3.63 (m, 1H, H-7a), 3.17 (m, 1H, H-5 *trans* to H-7a), 2.88 (m, 1H, H-5 *cis* to H-7a), 2.04 (m, 1H, H-6), 1.91 (m, 2H, H-6, H-7), 1.73 (m, 1H, H-7), 1.42 (s, 3H, CH₃), 1.32 (t, 3H, CH₃CH₂O); ¹³C NMR (100.1 MHz, CDCl₃): δ 172.5 (s), 137.8 (s), 128.9 (2d), 127.7 (2d), 127.6 (d), 99.2 (d, C-2), 70.6 (s, C-3), 70.2 (d, C-7a), 62.2 (t, CH₂O), 54.8 (d, C-1), 46.6 (t, C-5), 29.0 (t, C-7), 25.2 (t, C-6), 16.0 (q, CH₃), 14.0 (q, CH₃CH₂O); *m*/z 319 (10, M+1), 272 (40, M—NO₂), 245 (90, M—COOEt, 199 (100, M—COOEt—NO₂), 198 (80, M—COOEt—HNO₂), 184 (80, 199—CH₃). Anal. Calcd. for C₁₇H₂₂N₂O₄: C, 64.13; H, 6.97; N, 8.80; Found: C, 64.86; H, 6.74; N, 8.68.

(1S*,2S*,3R*,7aS*)-3-Benzoyl-1-nitro-2-phenylazabicy-

clo[**3.0**] **octane** (**3c**). Oil, 80% yield, after purification. R_f 0.15 (eluent: light petroleum–ethyl acetate 4:1). IR (neat) 3088, 3065, 1598, 1581, 1496, 754, 698 (Ph), 1686 (C=O), 1545, 1375 (NO₂); ¹H NMR (400 MHz, CDCl₃): δ 7.65 (bd, 2H, *o*-PhCO), 7.45 (bt, 1H, *p*-PhCO), 7.32 (bt, 2H, *m*-PhCO), 7.09 (m, 5H, Ph), 5.48 (dd, $J_1 = 7.8$ Hz, $J_2 = 5.3$ Hz, 1H, H-1), 5.32 (d, J = 7.3 Hz, 1H, H-3), 4.49 (m, 2H, H-2, H-7a), 3.42 (m, 1H, H-5), 2.81 (m, 1H, H-5), 2.06 (m, 3H, 2 H-6, H-7), 1.49 (m, 1H, H-7); ¹H NMR (400 MHz, CD₃OH): δ 8.05 (bd, 2H, *o*-PhCO), 7.70 (bd, 2H, *o*-Ph), 7.55 (bt, 1H, *p*-PhCO), 7.32 (bt, 2H, *m*-PhCO), 7.10 (m, 3H, Ph), 5.50 (t, J = 7.8 Hz, 9.5 Hz, 1H, H-1), 5.18 (d, J = 7.7 Hz, 1H, H-3), 4.51 (dd, $J_1 = 9.5$ Hz, $J_2 = 7.7$ Hz, 1H, H-2), 4.31 (dt, $J_1 = 9.5$ Hz, $J_2 = 7.7$ Hz, 1H, H-5), 2.89 (m, 1H, H-5), 2.30 (m, 3H, 2 H-6, H-7), 2.15 (m, 1H, H-7); ¹³C NMR (100.1



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MHz, CDCl₃): δ 198.2 (s), 136.8 (s), 135.1 (s), 133.0 (d, *p*-PhCO), 128.8 (2d, Ph), 128.5 (2d, *m*-PhCO), 128.1 (d, Ph), 127.9 (2d, Ph), 127.9 (2d, *o*-PhCO), 92.6 (d, C-1), 73.5 (s, C-3), 67.0 (d, C-7a), 55.6 (t, C-5), 53.3 (d, C-2), 28.2 (t, C-7), 27.4 (t, C-6); *m*/*z* 317 (2, M+1), 286 (20), 231 (95, M—PhCO), 184 (100, M—PhCO—HNO₂), 156 (45). Anal. Calcd. for $C_{20}H_{20}N_2O_3$: C, 71.41; H, 5.99; N, 8.33; Found: C, 71.02; H, 5.72; N, 8.40.

(1S,2S,3R,6R,7aS)-3-Acetyl-6-hydroxy-3-methyl-1-nitro-2phenylazabicyclo[3.3.0]octane (6a). Colorless solid, mp 118-119°C, 48% yield, after purification. Rf 0.10 (eluent: n-heptane: ethyl acetate 1:3). IR (CDCl₃) 3399 (OH), 3051, 3032, 1590, 1501, 734, 702 (Ph), 1706 (C=O), 1546, 1376 (NO₂); ¹H NMR (400 MHz, CDCl₃): δ 7.25 (m, 5H, Ph), 5.95 (dd, J_1 = 10.2 Hz, J_2 = 9.1 Hz, 1H, H-1), 4.51 (bq, J = 8.2 Hz, 1H, H-7a), 4.49 (b signal, 1H, H-6), 3.87 (d, J = 10.2 Hz, 1H, H-2), 3.13, 3.06 (part AB of an ABX system, $J_{AB} = 9.5$ Hz, J_{AX} = 3.3 Hz, $J_{\rm BX}$ = 1.2 Hz, 2H, 2 H-5), 2.12 (dd, J_1 = 8.2 Hz, $J_2 = 14.3$ Hz, 1H, H-7), 2.06 (bs, 1H, OH), 1.91 (s, 3H, CH₃CO), 1.65 (ddd, $J_1 = 14.3$ Hz, $J_2 = 7.9$ Hz, $J_3 = 5.3$ Hz, 1H, H-7), 1.56 (s, 3H, CH₃); ¹³C NMR (100.1 MHz, CDCl₃): δ 207.6 (s), 133.5 (s), 129.0 (2d), 128.7 (2d), 127.9 (d), 93.4 (d, C-1), 74.5 (s, C-3), 72.0 (d, C-6), 62.8 (d, C-7a), 56.2 (t, C-5), 55.9 (d, C-2), 36.9 (t, C-7), 27.3 (q, CH₃CO), 19.4 (q, CH₃); m/z 304 (<1, M^{+•}), 261 (56, M—CH₃CO), 215 (28, M-CH₃CO-NO₂), 214 (100, M-CH₃CO-HNO₂), 196 (25, 214—H₂O), 170 (10); Anal. Calcd. for C₁₆H₂₀N₂O₃: C, 63.14; H, 6.62; N, 9.20; Found: C, 63.27; H, 6.80; N, 9.09; $[\alpha]_D^{25} =$ -120.5 (*c* 0.36, CHCl₃).

(1S,2R,3R,6R,7aS)-3-Acetyl-6-hydroxy-3-methyl-2-nitro-1phenylazabicyclo[3.3.0]octane (7a). Semisolid material, 15% yield, after purification, R_f 0.23 (eluent: *n*-heptane: ethyl acetate 1:3). IR (CDCl₃) 3406 (OH), 3050, 3032, 1603, 1496, 731, 701 (Ph), 1713 (C=O), 1549, 1359 (NO₂); ¹H NMR (400 MHz, CDCl₃): δ 7.35-7.25 (m, 3H, Ph), 7.15 (bd, 2H, Ph), 6.10 (d, J = 8.1 Hz, 1H, H-2), 4.57 (b signal, 1H, H-6), 3.83 (dd, $J_1 = 8.1$ Hz, $J_2 = 10.6$ Hz, 1H, H-1), 3.39, 3.37 (m + dd, $J_1 = 4.0$ Hz, $J_2 = 9.9$ Hz, 2H, H-7a, H-5), 3.05 (bd, J =9.9 Hz, 1H, H-5), 2.42 (s, 3H, CH₃CO), 2.18 (ddd, $J_1 = 3.3$ Hz, $J_2 = 5.9$ Hz, $J_3 = 14.6$ Hz, 1H, H-7), 2.06 (s + ddt, $J_1 =$ 14.6 Hz, $J_2 = 7.5$ Hz, $J_3 = 1.2$ Hz, 2H, OH + H-7), 1.39 (s, 3H, CH₃); ¹H NMR (400 MHz, CD₃OH): δ 7.30 (m, 5H, Ph), 6.00 (d, J = 8.8 Hz, 1H, H-2), 4.55 (b signal, 1H, H-6), 3.82 (dd, $J_1 = 8.8$ Hz, $J_2 = 9.9$ Hz, 1H, H-1), 3.46 (ddd, 1H, H-7a), 3.36 (dd, $J_1 = 4.6$ Hz, $J_2 = 10.1$ Hz, 1H, H-5), 2.99 (d, J = 10.1 Hz, 1H, H-5), 2.43 (s, 3H, CH_3CO), 2.14 (ddd, 1H, H-7), 1.91 (ddt, 1H, H-7), 1.34 (s, 3H, CH₃); ¹³C NMR (100.1 MHz, CDCl₃): δ 207.0 (s), 137.2 (s), 129.1 (2d), 127.8 (2d), 127.7 (d), 98.5 (d, C-2), 76.1 (s, C-3), 72.3 (d, C-6), 69.1 (d, C-7a), 57.1 (d, C-1), 55.1 (t, C-5), 39.7 (t, C-7), 24.2 (q, CH₃CO), 16.2 (q, CH₃); *m*/*z* 304 (<1, M^{+•}), 261 (58, M— CH₃CO), 215 (100, M-CH₃CO-NO₂), 214 (18, M-CH₃CO-HNO₂), 200 (28), 170 (15). Anal. Calcd. for $C_{16}H_{20}N_2O_3{:}$ C, 63.14; H, 6.62; N, 9.20; Found: C, 63.32; H, 6.85; N, 9.02; $[\alpha]_D^{25} = +71.8$ (*c* 0.87, CHCl₃).

(1R,2S,3S,6R,7aR)-3-Acetyl-6-hydroxy-3-methyl-2-nitro-1phenylazabicyclo[3.3.0]octane (8a). Colorless solid, mp 166– 167°C, 15% yield, after purification, R_f 0.27 (eluent: *n*-heptane: ethyl acetate 1:3). IR (nujol) 3181 (OH), 2724, 2671 (CH), 1713 (C=O), 1602, 1493, 701 (Ph), 1549, 1359 (NO₂); ¹H NMR (400 MHz, CDCl₃): δ 7.36 (m, 3H, Ph), 7.25 (bd, 2H, Ph), 6.10 (d, J = 8.4 Hz, 1H, H-2), 4.59 (b signal, 1H, H-6), 4.18 (dd, $J_1 = 8.4$ Hz, $J_2 = 10.5$ Hz, 1H, H-1), 3.35 (dd, $J_1 = 9.0$ Hz, $J_2 = 6.4$ Hz, 1H, H-5), 3.22 (dd, $J_1 = 9.0$ Hz, $J_2 = 7.3$ Hz, 1H, H-5), 3.20 (ddd, $J_1 = 2.9$ Hz, $J_2 = 7.7$ Hz, $J_3 = 10.5$ Hz, 1H, H-7a), 2.34 (s + m, 4H, CH₃CO, H-7), 1.84 (bd, 1H, OH), 1.78 (ddd, $J_1 = 2.9$ Hz, $J_2 = 5.1$ Hz, $J_3 = 13.7$ Hz, 1H, H-7), 1.38 (s, 3H, CH₃); ¹³C NMR (100.1 MHz, CDCl₃): δ 207.0 (s), 137.4 (s), 129.0 (2d), 127.8 (2d), 127.7 (d), 98.0 (d, C-2), 77.2 (s, C-3), 72.3 (d, C-6), 69.8 (d, C-7a), 57.1 (d, C-1), 54.4 (t, C-5), 38.0 (t, C-7), 24.1 (q, CH₃CO), 16.2 (q, CH₃); m/z 304 (<1, $M^{+\bullet}$), 261 (56, M—CH₃CO), 215 (30, M—CH₃CO—NO₂), 214 (100, M—CH₃CO—HNO₂), 196 (32, 214—H₂O), 170 (12). Anal. Calcd. for C₁₆H₂₀N₂O₃: C, 63.14; H, 6.62; N, 9.20; Found: C, 63.22; H, 6.74; N, 9.13; $[\alpha]_D^{25} = -83.3$ (*c* 0.09, CHCl₃).

(1'S*,2'R*,3'S*,7'aS*)-2'-Nitro-1'-phenylspiro[cyclohexanone-6, 3'-pyrrolizidine (10). Colorless oil, 70% yield, after purification. IR (neat) 3063, 3029, 1603, 1497, 757, 701 (Ph), 1713 (CO), 1545, 1361 (NO₂); ¹H NMR (400 MHz, CDCl₃): δ 7.30 (m, 5H, Ph), 6.16 (d, J = 7.7 Hz, 1H, H-2'), 3.81 (dd, $J_1 = 10.6$ Hz, J_2 = 7.7 Hz, 1H, H-1'), 3.28–3.15 (m, 3H, H-2, H-5', H-7'a) [The three overlapping signals are as follows: 3.20 (dt, $J_1 = J_2 = 14.0$ Hz, $J_3 = 6.0$ Hz, 1H, H-2), 3.20 (m, 2H, H-5'), 3.18 (m, 1H, H-7'a)], 3.12 (dt, 1H, H-5'), 2.39 (bd, J = 14.0 Hz, 1H, H-2), 2.20– 2.05 (m, 3H, H-5, H-6', H-7'), 2.00-1.77 (m, 4H, H-3, H-4, H-6', H-7'), 1.77–1.58 (m + dt, 2H, H-3, H-5); ¹³C NMR (100.1 MHz, CDCl₃): δ 207.8 (s), 138.2 (s), 128.9 (2d), 128.1 (2d), 127.4 (d), 98.3 (d, C-2'), 76.9 (s, C-3'), 70.7 (d, C-7'a), 56.8 (d, C-1'), 45.0 (t, C-5'), 37.9 (t, C-2), 30.9 (t, C-5), 27.0 (t, C-7'), 26.9 (t, C-3), 23.4 (t, C-6'), 22.2 (t, C-4); m/z 315 (10, M+1), 268 (78, M+1-NO2), 240 (45, 268-CO), 171 (100), 143 (30), 129 (92), 70 (65). Anal. Calcd. for C18H22N2O3: C, 68.77; H, 7.05; N, 8.91; Found: C, 68.91; H, 7.25; N, 8.75.

(1S*,5S*,6R*,7R)-1-Hydroxy-7-nitro-6-phenylbicyclo[3.2.1] octan-8-one (11). Semisolid material, 73% yield, after purification, $R_{\rm f}$ 0.40 (eluent: light petroleum-ethyl acetate 4:1). IR (CCl₄) 3454 (OH), 3050, 3030, 1498, 788, 754, 700 (Ph), 1764 (C=O), 1549, 1370 (NO₂); ¹H NMR (400 MHz, CDCl₃): δ 7.30 (m, 3H, Ph), 7.15 (bd, 2H, Ph), 4.81 (d, J = 5.8 Hz, 1H, H-7), 4.18 (d, J = 5.8 Hz, 1H, H-6), 3.36 (bs, 1H, OH), 2.78 (dd, $J_1 = 4.4$ Hz, $J_2 = 1.8$ Hz, 1H, H-5), 2.35 (m, 2H, CH₂), 2.11 (m, 1H, ring CH), 2.00 (s, 2H, CH₂), 1.75 (m, 1H, ring CH); 13C NMR (100.1 MHz, CDCl₃): δ 212.5 (s), 142.3 (s), 129.4 (2d), 127.8 (d), 126.7 (2d), 93.6 (d, C-7), 81.6 (s, C-1), 51.6 (d, C-5), 43.9 (d, C-6), 39.8 (t, C-2), 36.0 (t, C-4), 17.9 (t, C-3); m/z 261 (11, M^{+•}), 229 (20, M—NO—H₂), 197 $(10, M-NO_2-H_2O), 169 (100, 197-28), 141 (48, 169-28).$ Anal. Calcd. for C₁₄H₁₅NO₄: C, 64.36; H, 5.79; N, 5.36; Found: C, 64.40; H, 5.87; N, 5.40.

(1S*,5S*,6R*,7S)]-1-Hydroxy-7-nitro-6-phenylbicyclo[3.2.1] octan-8-one (12). Mp 146–147°C, from cyclohexane, 7% yield, after purification. R_f 0.30 (eluent: light petroleum–ethyl acetate 4:1). IR (nujol) 3425 (OH), 3050, 3030, 1496, 743, 721, 695 (Ph), 1752 (C=O), 1553 (NO₂); ¹H NMR (400 MHz, CDCl₃): δ 7.28 (m, 3H, Ph), 7.16 (bd, 2H, Ph), 5.29 (d, J = 9.5 Hz, 1H, H-7), 3.92 (d, J = 9.5 Hz, 1H, H-6), 3.22 (bs, 1H, OH), 3.05 (b signal, $W_H = 9.2$ Hz, 1H, H-5), 2.41 (m, 1H), 2.20– 2.05 (m, 3H, ring CH), 2.02–1.83 (m, 2H); ¹³C NMR (100.1 MHz, CDCl₃): δ 213.9 (s), 136.3 (s), 128.7 (2d), 128.5 (2d), 127.8 (d), 91.0 (d, C-7), 80.0 (s, C-1), 47.1 (d, C-5), 44.3 (d, C-6), 41.2 (t, C-2), 35.9 (t, C-4), 17.6 (t, C-3). Anal. Calcd. for $C_{14}H_{15}NO_4$: C, 64.36; H, 5.79; N, 5.36; Found: C, 64.42; H, 5.93; N, 5.50.

Crystal structure determinations. Diffraction data for the structures reported were collected at room temperature on a Nonius DIP-1030H system (Mo-K α radiation, $\lambda = 0.71073$ Å). Both the structures were solved by direct methods and refined by the full-matrix least-squares method based on F^2 with all observed reflections [8]. The calculations were performed using the WinGX System, Ver 1.80.05 [9].

Crystal data for 6a. C_{16} ·H₂₀·N₂·O₄, fw = 304.34 g/mol; Tetragonal, P 4₁, a = 13.640(3), c = 8.299(2) Å, V = 1544.0(6) Å³, Z = 4, D_{calcd} = 1.309 g/cm³, μ (Mo-K α) = 0.095 mm⁻¹, F(000) = 648, θ = 24.71°. Final R_1 = 0.0426, wR_2 = 0.0996, GOF = 0.850 for 132 parameters and 1370 unique reflections, of which 815 with $I > 2\sigma(I)$, residuals in ΔF map 0.186, -0.181 e. Å⁻³.

Crystal data for 12. C_{14} ·H₁₅·NO₄, fw = 261.27 g/mol; Monoclinic, P_{21} , a = 8.323(2), b = 6.309(2), c = 12.535(3)Å, $\beta = 104.31(3)^{\circ}$, V = 637.8(3) Å³, Z = 2, $D_{calcd} = 1.361$ g/ cm³, μ (Mo-K α) = 0.100 mm⁻¹, F(000) = 276, $\theta = 24.69^{\circ}$. Final $R_1 = 0.0341$, $wR_2 = 0.0890$, GOF = 0.889 for 173 parameters and 1107 unique reflections, of which 717 with $I > 2\sigma(I)$, residuals in ΔF map 0.101, -0.156 e. Å⁻³.

CONCLUSION

Although [3+2]cycloadditions between 1,3-azomethine ylides and activated olefins are known, the easy access to these polyfunctionalized pyrrolizidines is interesting, in particular, as far as the formation of the nitro spiro pyrrolizidine is concerned. In fact, a similar spiro pyrrolizidine of terpenoid nature, the (+)-nitropolyzonamine [10], extracted from a millipede, has been studied as a potential allomone to deter predators [11].

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